

100

BIO BASIC Worldwide



For more information on pricing, complete product line or to locate a Point of Sales near you, please visit our website or contact one of our Customer Service Representatives.

Email	\square	order@biobasic.com		
Phone	5	1	(905)	474-4493
Toll Free	5	1	(800)	313-7224
Fax		1	(905)	474-5794



9K-006-0023s (1 prep) 9K-006-0023 (10 prep) 9K-006-0026 (30 prep)

QF 24 V5 V2 October 2021

For Research Use Only

Plasmid DNA Extraction Maxiprep Kit

Table of Contents

Introduction	02
Specifications	02
Kit Contents	02
Important Notes	03
Additional Materials Required	03
Brief Procedure	04
General Protocol	06
Troubleshooting Guide	06

Plasmid DNA Extraction Maxiprep Kit

Introduction

Feldan Plasmid DNA Extraction Maxiprep Kit is an excellent tool offering a rapid and economic method to purify plasmid DNA from bacteria cultures. This technology is based on alkaline lysis and purification by Anion-exchange chromatography. Compared with other harmful and time-consuming procedures such as phenol / chloroform extraction and ethanol precipitation, Feldan Plasmid DNA Extraction Kit shortens the handling time to about 2 hours. The high quality plasmid DNA can be used directly for any downstream applications.

Specifications

Sample Size	Yield	Handling Time
100~250 mL of bacteria culture for high copy plasmids	up to 500 µg for	About 2 hours
200~400 mL of bacteria culture for low copy plasmids	high-copy plasmids	

Kit Contents

Component	9K-006-0023s, 1 Preps	9K-006-0023, 10 Preps	9K-006-0026 30 Preps
MAXI 1 Resuspension Solution	11ml	110ml	3x110ml
MAXI 2 Cell Lysis Solution	11ml	110ml	3x110ml
MAXI 3 Neutralization Solution	11ml	110ml	3x110ml
MAXI 4 Equilibration Solution	13.5ml	135ml	3x135ml
MAXI 5 Washing Solution	55ml	2x275ml	6x275ml
MAXI 6 Elution Solution	13.5ml	135ml	3x135ml
RNase A (50mg/ml)	22µl	220µl	3x220µl
MAXI Column	1 pc	10 pcs	30 pcs
Protocol	1	1	1

Important Notes

- 1 Solutions provided in this kit contain irritants, wear gloves and lab coat when handling.
- 2. Briefly spin RNase A tube to remove drops from the inside of the lid. Transfer tube content into MAXI 1 Resuspension Solution bottle. Add 250μL of MAXI 1 Resuspension Solution into RNase A tube, rinse tube inside and transfer back into MAXI 1 resuspension Solution bottle. Store at 4°C.
- 3. Check MAXI 2 Cell Lysis Solution before use. Warm MAXI 2 Cell Lysis Solution at 37°C if any pre-cipitation formed. Prevent vigorous shaking of the MAXI 2 Cell Lysis Solution.
- 4. To avoid acidification of MAXI 2 Cell Lysis Solution from CO2 in the air, close the bottle immediately after use.





Additional Material Required

- 1. 50 mL centrifuge tube.
- 2. Isopropanol
- 3. 70 % Ethanol

General Protocol

- 1. Harvest the bacterial culture by centrifugation at 6,000 x g for 15 minutes.
- 2. Add 10 ml of MAXI 1 Resuspension Solution (RNase A added) and resuspend the cell pellet by vortexing or pipetting.
- 3. Add 10 ml of MAXI 2 Cell Lysis Solution and mix gently by inverting the tube 10 times. Do not vortex to avoid shearing of genomic DNA.
- 4. Incubate for 3 minutes at room temperature until lysate clears.
- 5. Add 10 mL of MAXI 3 Neutralization Solution and mix immediately by inverting the tube 10 times. Do not vortex to avoid shearing of genomic DNA.
- 6. Centrifuge at 15,000 x g for 20 minutes at 4 °C.
- 7. Transfer supernatant from step 6 in a new tube. Centrifuge at 15,000 x g for 20 minutes at 4°C.
- Place a MAXI Column into a 50 mL centrifuge tube, add 10 mL of MAXI 4 Equilibration Solution to equilibrate the MAXI Column and allow the column to empty by gravity flow.
- 9. Transfer the supernatant from step 7 to the equilibrated MAXI Column, and allow the column to empty by gravity flow. Discard the filtrate.
- 10. Add 25 ml of MAXI 5 Washing Solution to wash the MAXI Column and allow the column to empty by gravity flow. Discard the filtrate.
- 11. Repeat step 10.
- 12. Place the MAXI Column into a clean 50 mL centrifuge tube (not provided) and add 12 mL of MAXI 6 Elution Solution to elute DNA by gravity flow.
- 13. Precipitate DNA by adding 9 mL of isopropanol to the eluted DNA from Step 12.
- 14. Mix gently and centrifuge at 20,000 g for 30 minutes at 4 °C.
- 15. Carefully remove the supernatant and wash the DNA pellet with 5 mL of room temperature 70% ethanol.
- 16. Centrifuge at 20,000 g for 10 minutes at 4°C.
- 17. Carefully remove the supernatant and air-dry the DNA pellet for 10 minutes.
- 18. Dissolve the DNA pellet in a suitable volume of 10mM tris pH 8.5 or ddH₂O

Troubleshooting

Low Yield

Bacterial cells were not lysed completely.

- Too many bacterial cells were used.
- After MAXI 3 Neutralization Solution addition, break up the precipitate by inverting.
- DNA pellet was lost after precipitation.
- DNA pellet was insufficiently redissolved.

Purified DNA doesn't perform well in downstream application.

RNA contamination

- Make sure that RNase A was has been added in MAXI 1 Resuspension Solution when first using. If RNase A added in MAXI 1 Resuspension Solution has expired, add additional RNase A.
- Too many bacterial cells were used, reduce the sample volume.
- Elution buffer contains EDTA.

Genomic DNA contamination

- Do not use overgrown bacterial culture.
- During MAXI 2 Cell Lysis Solution and MAXI 3 Neutralization Solution addition, mix gently to prevent genomic DNA shearing.
- Lysis time was too long (over 5 minutes).

Too much salt residual in DNA pellet

• Wash the DNA pellet twice with 70% ethanol