

Plasmid DNA Extraction

BIO BASIC Worldwide



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MAXI Prep Kit Bs4654 (4 preps) Bs466 (20 preps) Version 5.0A Rev 31.8.2015

For Research Use Only

Introduction

Bio Basic Plasmid DNA Extraction Maxiprep Kit is an excellent tool offering a rapid and economic method to purify plasmid DNA from bacterial cultures. This technology is based on alkaline lysis and purification by Anion-exchange chromatography. Compared with other harmful and time-consuming procedures such as phenol / chloroform extraction and ethanol precipitation, Bio Basic Plasmid DNA Extraction Kit shortens the handling time to about 2 hours. The high quality plasmid DNA can be used directly for any downstream application.

Specification

| Sample Size | Yield | Handling Time |
|--|--|------------------|
| 100-250mL of bacterial culture for high copy plasmids 200-400 mL of bacterial culture for low copy plasmids | Up to 500 µg for high copy- plasmids | About 2 hours |

- 16. Centrifuge at $20,000 \times g$ for 10 minutes at $4^{\circ}C$.
- 17. Carefully remove the supernatant and air-dry the DNA pellet for 10 minutes.
- 18. Dissolve the DNA pellet in a suitable volume of 10mM tris pH8.5 or ddH_2O .

Troubleshooting

Low yield

Bacterial cells were not lysed completely.

- Too many bacterial cells were used.
- After MAXI 3 Neutralization Solution, break up the precipitate by inverting the tube several times.
- DNA pellet was lost after precipitation.
- DNA pellet was insufficiently redissolved.

Purified DNA doesn't perform well in downstream application

RNA contamination

- Make sure that RNase A has been added in MAXI 1 Resuspension Solution when first using. If RNase A added in MAXI 1 Resuspension Solution has expired, add additional RNase A.
- Too many bacterial cells were used, reduce the sample volume.
- Elution buffer contains EDTA.

Genomic DNA contamination

- Do not use overgrown bacteria culture.
- During MAXI 2 Cell Lysis Solution and MAXI 3 Neutralization Solution addition, mix gently to prevent genomic DNA shearing.
- Lysis time was too long (over 5 minutes).

Too much salt residual in DNA pellet

• Wash the DNA pellet twice with 70% ethanol.

- 5. Add 10 mL of MAXI 3 *Neutralization Solution* and mix immediately by inverting the tube 10 times. Do not vortex to avoid shearing of genomic DNA.
- 6. Centrifuge at 15,000 x g for 20 minutes at 4°C.
- 7. Transfer supernatant from step 6 in a new tube. Centrifuge at 15,000 x g for 20 minutes at 4°C.
- Place a MAXI Column into a 50 mL centrifuge tube, add 10 mL of MAXI 4 Equilibration Solution to equilibrate the MAXI Column and allow the column to empty by gravity flow. Discard the filtrate
- 9. Transfer the supernatant from step 7 to the equilibrated MAXI Column, and allow column to empty by gravity flow. Discard the filtrate.
- 10. Add 25 mL of MAXI 5 Washing Solution to wash the MAXI Column and allow the column to empty by gravity flow. Discard the filtrate.
- 11. Repeat Step 10.
- 12. Place MAXI Column into a clean 50 mL centrifuge tube (not provided) and add 12 mL of MAXI 6 Elution Solution to elute DNA by gravity flow.
- 13. Precipitate DNA by adding 9mL of isopropanol to the eluted DNA from Step 12.
- 14. Mix gently and centrifuge at 20,000 x g for 20 to 30 minutes at 4°C.
- 15. Carefully remove the supernatant and wash the DNA pellet with 5mL of room temperature 70% ethanol.

Content

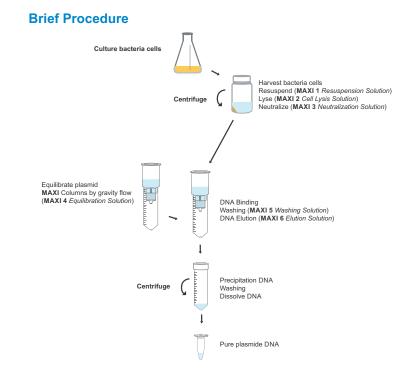
| Component | BS4654 (4 preps) | BS466 (20 preps) |
|--------------------------------------|---------------------|---------------------|
| MAXI 1 Resuspension Solution | 55mL | 220 mL |
| MAXI 2 Cell Lysis Solution | 55 mL | 220 mL |
| MAXI 3 Neutralization Solution | 55 mL | 220 mL |
| MAXI 4 Equilibration Solution | 54 mL | 270 mL |
| MAXI 5 Washing Solution | 4X55 mL | 4x275 mL |
| MAXI 6 Elution Solution | 54 mL | 270 mL |
| RNase A (50mg/mL) | 88 µL | 440 µL |
| MAXI Column | 4 pcs | 20 pcs |
| User Manual | 1 | 1 |

Additional Materials Required

- 1. 50 mL centrifuge tube.
- 2. Isopropanol
- 3. 70% Ethanol

Important Notes

- 1. Solutions provided in this kit contain irritants, wear gloves and lab coat when handling.
- Briefly spin RNase A tube to remove drops from the inside of the lid. Transfer tube contents into MAXI 1 *Resuspension Solution* bottle. Add 250 µL of MAXI 1 *Resuspension Solution* into RNase A tube, rinse tube inside and transfer back into MAXI 1 *Resuspension Solution* bottle. Store at 4°C.
- Check MAXI 2 Cell Lysis Solution before use. Warm MAXI 2 Cell Lysis Solution at 37°C if any precipitation formed. Prevent vigorous shaking of the MAXI 2 Cell Lysis Solution.
- 4. To avoid acidification of MAXI 2 *Cell Lysis Solution* from CO₂ in the air, close the bottle immediately after use.



General Protocol

- 1. Harvest the bacterial culture by centrifugation at 6,000 x g for 15 minutes.
- 2. Add 10 mL of MAXI 1 *Resuspension Solution* (RNase A added) and resuspend the cell pellet by vortexing or pippetting.
- 3. Add 10 mL of MAXI 2 *Cell Lysis Solution* and mix gently by inverting the tube 10 times. Do not vortex to avoid shearing of genomic DNA.
- 4. Incubate for 3 minutes at room temperature until lysate clears.

EZ Spin Column Handbook

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