



## **PRODUCT INFORMATION**

### **DNA, RNA-EZ Q2 (DNase, RNase-Be-Gone) Solution**

#### ***Product information for RT4201:***

#### **Component:**

<b>Component</b>	<b>RT4201, 50 ml</b>	<b>RT4201, 5 × 50 ml</b>
DNase, RNase-Be-Gone(surface)	50 ml	5 × 50 ml
Protocol	1	1

#### **Storage:**

Transportation and storage at room temperature.

#### **Features**

1. Effectively inactivates up to 100 µg of surface-contaminated RNases such as RNase T1, RNase H, BAL31, S1, Mung bean nuclease etc. and DNases in 10 minutes.
2. Faster than using DEPC.
3. Non-toxic.
4. Economical.
5. Simple: Simply dilute DNase & RNase-Be-Gone solution with water in a ratio of 1:1000 (v/v) and wash surfaces of glasswares, plasticwares and instruments. keep for 10 minutes at room temperature.
6. Can be used for making RNase-free water. For making RNase-free water add 0.1% of DNase & RNase & RNase-Be-Gone solution to dd-water and keep for at least 24 hours at room temperature.

#### **Introduction**

The Surface DNase & RNase-Be-Gone is a complex solution designed for removing >80% surface-RNase from tubes, tips, pipettes, glasswares and instruments surface

#### **Protocol**

##### **Making RNase-free water.**

1. Add 0.1% of RNase-Be-Gone B solution to dd-water and keep for 24 hours or longer at room temperature.
2. The RNase-free water is appropriate for making lysis buffer or other solution.

##### **Clean the operating desktop.**

1. Dilute RNase-Be-Gone solution with water in a ratio of 1:10 (v/v). Mix well and spray it over the operating desktop.



2. Incubate at room temperature for 5 minutes or longer. Wipe the residual solution with absorbent paper.
3. Wash the operating desktop once with the diluted RNase-Be-Gone solution in ratio of 1:1000 (v/v). Wipe the residual solution with absorbent paper to dry the operating desktop.

Note: We recommend RNase-Be-Gone solution for directly using to clean the operating desktop serious polluted RNases.

#### **Clean the instruments.**

1. Firstly, wipe the surface of instruments with diluted RNase-Be-Gone solution in ratio of 1:10 (v/v).
2. And then wipe the surface of instruments once with diluted RNase-Be-Gone solution in ratio of 1:1000 (v/v).
3. The RNase-Be-Gone solution has corrosives to metallic instruments, protects is for use time operation should be less 5 minutes.

Note: We recommend RNase-Be-Gone solution for directly using to clean the instruments serious polluted RNases.

#### **Clean glasswares and plasticwares.**

1. Firstly, glasswares and plasticwares should soak for 5 minutes in diluted RNase-Be-Gone solution in ratio of 1:10 (v/v) or 1:100 (v/v).
2. And then wash the glasswares and plasticwares once with diluted RNase-Be-Gone solution in ratio of 1:1000 (v/v) or 1:10000 (v/v).
3. Dry the glasswares and plasticwares completely.

#### **Clean pipettes**

1. Pipette dismounting should be performed according to manufacturer recommended procedures.
2. The component of the pipette should soak for 1 minute in diluted RNase-Be-Gone solution in ratio of 1:10 (v/v) or 1:100 (v/v).
3. And then wash it once with diluted RNase-Be-Gone solution in ratio of 1:1000 (v/v) or 1:10000 (v/v).
4. Dry the pipette completely.

#### **Clean pipette tips and microcentrifuge tubes**

1. Firstly, pipette tips and microcentrifuge tubes should soak for 5 minutes in diluted RNase-Be-Gone solution in ratio of 1:1000 (v/v).
2. And then wash the pipette tips and microcentrifuge tubes twice with diluted RNase-Be-Gone solution in ratio of 1:1000 (v/v) or 1:10000 (v/v).
3. Dry the pipette tips and microcentrifuge tubes completely.