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96-Well Plate Plant Genomic
DNA Mini-Preps Kit

BS8361
QF 24 TV4
CV1 2020

For Research Use Only

96-Well Plate Plant Genomic DNA Mini-Preps Kit Code: BS8361 (2 Plates)

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Description

The kit provides a rapid and convenient high throughput technique for mini preparation of high quality genomic DNA from various plants species. DNA of plant lysate is selectively absorbed on each column of the plate and other impurities such as proteins, salts are eliminated from the column. Each column can absorb up to 20 µg of DNA. Purified plant genomic DNA is 20-40 kb, and is suitable for downstream applications such as PCR, Real-time RT PCR, Southern blotting, Microsatellite analysis, AFLP, RFLP, and RAPD.

Features

- Fast and Simple. Using a rapid spin and high throughput format, the entire procedure takes less than 2 hours.
- High purity of DNA. OD_{260}/OD_{280} of purified DNA is generally >1.8.
- Compatible with many downstream applications such as PCR, restriction digestions, real-time PCR, multiplex PCR, RAPD, RFLP, AFLP, Southern Blotting and microsatellite analysis.
- Suitable for a wide variety of plant species and tissue types including some very recalcitrant specimens.

Other Kits Available

EZ-10 Spin Column Plasmid DNA Mini-Preps Kit

- BS413 (50 Preps)
- BS414 (100 Preps)
- BS614 (250 Preps)

EZ-10 Spin Column PCR Products Purification Kit

- BS363 (50 Preps)
- BS364 (100 Preps)
- BS664 (250 Preps)

EZ-10 Spin Column DNA Gel Extraction Kit

- BS353 (50 Preps)
- BS354 (100 Preps)
- BS654 (250 Preps)



PRODUCTS ARE FOR SCIENTIFIC RESEARCH ONLY
NOT INTENDED FOR HUMAN OR ANIMAL USE

10. To elute, place a 96 Well Storage Plate on top of a Deep Well Collection Plate, and then place the EZ-10 96 Well Binding Plate on the top of a 96 Well Storage Plate. Add 50 µl TE Buffer directly onto the center part of the EZ-10 96 Well Binding Plate membrane. Incubate at room temperature for 1 minute, and then centrifuge for 1 minute at 5,000 x g (6,000 rpm) to elute the DNA.

NOTE 1: Warm the TE Buffer to 60°C will increase the elution efficiency.

NOTE 2: Elution with more than 50 µl (e.g. 200 µl) increases the DNA yield, but the concentration will be lower.

NOTE 3: For maximum DNA yield, repeat elution once as described in this step.

NOTE 4: For maximum DNA concentration, use the eluate in the microcentrifuge tube for the second elution step.

Kit Contents

Components	BS8361 (2 Plates)
Universal Buffer PCB	130 ml
Universal Buffer BD	50 ml
Universal PW Solution	72 ml
Universal Wash Solution	30 ml
TE Buffer (pH 8.0)	40 ml
EZ-10 96 Well Binding Plate	2
Deep Well Collection Plate	6
96 Well Storage Plate	2
Sealing film	8
Protocol	1

NOTE 1: Universal Buffer BD contains chaotropic salt; avoid contact with skin and eyes.

NOTE 2: Universal PW Solution and Universal Wash Solution are supplied as concentrates. Add 48 ml isopropanol to 72 ml Universal PW Solution and 90 ml ethanol (96-100%) for 30 ml Universal Wash Solution before use to obtain a working solution.

Storage

96 Well Plates and all buffers should be stored dry, at room temperature (15-25°C). Kit is stable for 1 year under these conditions.

Materials Supplied by User

- Microcentrifuge capable of at least 6,000 × g
- Pipettes and pipette tips
- Vortexer
- Chloroform
- Isopropanol
- Ethanol (96-100%)
- RNase A (20 mg/ml, Optional for RNA-free DNA)
- Water bath for heating at 65°C
- β-mercaptoethanol

Procedures

1. Grind 100 mg fresh plant tissue (or 20 mg dry plant tissue) to fine powder in liquid nitrogen; transfer the powder to a Deep Well Collection Plate.

2. Add 600 µl Buffer PCB and 12 µl of β-mercaptoethanol to the sample. Seal the Deep Well Collection Plate with sealing film, and mix thoroughly by vortexing. Incubate at 65°C for 25 minutes.

NOTE 2: If RNA-free genomic DNA is required, add 20 µl RNase A (20 mg/ml), mix by vortexing, and incubate for 2 minutes at room temperature before continuing with step 3.

3. Add 0.6 ml of chloroform to each well. Seal the Deep Well Collection Plate with sealing film, and mix well by vortexing. Centrifuge at 5,700 × g for 2 minutes. Carefully transfer the supernatant (400 µl) to a new clean Deep Well Collection Plate.

4. Add 200 µl Universal Buffer BD. Seal the Deep Well Collection Plate with sealing film, and mix thoroughly by vortexing.

NOTE: If a gelatinous material appears at this step, incubate at 70°C for 10 minutes.

5. Add 200 µl ethanol (96-100%). Seal the Deep Well Collection Plate with sealing film, and mix thoroughly by vortexing.

NOTE: If a gelatinous material appears at this step, vigorously shaking or vortexing is recommended.

6. Transfer the mixture from step 5 (including any precipitate) into EZ-10 96 Well Binding Plate placed in a new Deep Well Collection Plate. Centrifuge at 5,000 × g (6,000 rpm) for 1 minute. Discard the flow-through.

7. Add 500 µl Universal PW Solution, and centrifuge for 1 minute at 5,000 × g (6,000 rpm). Discard the flow-through.

NOTE: Check the label to ensure PW Solution was diluted with isopropanol.

8. Add 500 µl Universal Wash Solution, and centrifuge for 1 minute at 5,000 × g (6,000 rpm). Discard the flow-through.

NOTE: Check the label to ensure Wash Solution was diluted with ethanol.

9. Place the empty EZ-10 96 Well Binding Plate in the Deep Well Collection Plate and centrifuge for an additional 2 minutes at 5,000 × g (6,000 rpm) to dry the EZ-10 96 Well Binding Plate membrane. Discard flow-through.

NOTE: It is important to dry the membrane of the EZ-10 96 Well Binding Plate, since residual ethanol may interfere with subsequent reactions. This centrifugation step ensures that no residual ethanol will be carried over during the following elution.